

# Early Development and Allometric Growth in Laboratory-Reared European Chub *Leuciscus cephalus* (Linnaeus, 1758)

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#### **Abstract**

The morphological development and allometric growth patterns in European chub *Leuciscus cephalus* (L.) (cyprinid fish) were studied under intensive rearing conditions from hatch to day 30. Based on the external morphology, twelve different stages during the early development of chub were identified (ES8<sub>b</sub>-JS1<sub>a</sub>; 6.68±0.20-20.84±2.15 mm TL). The obtained results revealed that the majority of all allometric changes from the inflection point occurred across a wide range of body lengths. The replacement of temporary organs (e.g. disappearance of finfold) at approximately 21 mm TL did not coincide with a full reduction of relative growth. This growth pattern probably reflects he absence of abrupt changes in the behaviour and habitat of young chubs.

Keywords: Leuciscus cephalus, early ontogeny, morphologic development, allometry.

# Introduction

Leuciscus cephalus (L.) is a rheophilic cyprinid widely distributed throughout Europe (Banarescu, 1992). However, the abundance of this species, as well as other from genus Leuciscus, has decreased in some natural populations because of human impact and partly by interspecific hybridization (Durand et al., 2000; Freyhof et al., 2005; Bolland et al., 2008; Kirtiklis et al., 2013; Nowosad et al., 2014). European chub is a fractional, late-spring spawner. Spawning grounds are characterized by 20-50 cm s<sup>-1</sup> current velocity, shallow depth (0.1-0.8 m) and a stony bottom, where most eggs adhered (Mann, 1996; Friedrich et al., 2003; Telechtea et al., 2009). The free embryos developed early photophobia and tended to congregate between pebbles. Young chubs shows schooling behavior and prefer those parts of streams and rivers where water currents are minimal (Economou et al., 1991). This species is considered a natural and essential component of many freshwater ecosystems, and they also have some practical meaning, e. g. considerable value in sport fishing, bioindicator of pollution (Hajkova et al., 2007) as well as a model species used for toxicity tests (Gomułka et al., 2011).

Thus far, studies on larval chub development have been conducted by Kryzhanovskij (1949);

Koblickaya (1981); Economou et al. (1990); Çalta (2000) and Kupren et al. (2008). None of these studies have described the morphological changes during the earliest stages of development in great detail. The transition from larval to juvenile development (metamorphosis) is associated with a shift in allometric growth or shape coupled with an abrupt or gradual changes in morphological characters (e.g. disappearance of finfold, acquisition of the adult complement of fin rays or squamation (Urho, 2002; Nikolioudakis et al., 2010). From an ecological point of view its most significant aspect is the occurrence of a niche and habitat shift of larvae (Urho, 2002; Kupren et al., 2014a). The objective of the present study was to provide biological knowledge on the morphological and functional development of European chub free embryos, larvae and early juvenile reared under intensive conditions, with a focus on the age and size at transformation. An understanding of the morphological development and allometric growth patterns provides insight into possible functional trends and environmental preferences of different developmental stages and thus is crucial for optimization of production in aquaculture (Koumoundouros et al., 1999; Gisbert et al., 2002; Choo and Liew, 2006). Moreover they may be used for determining the quality of chub juveniles in restocking programs.

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# **Materials and Methods**

European chub free embryos were obtained by the controlled reproduction of wild spawners caught in Lake Mosag (northeast Poland) at the beginning of the reproductive season. The reproduction was performed according to the method described by Krejszeff et al. (2008) with the application of the double hormonal treatment with Ovopel as was described by Targonska and Kucharczyk (2011) for goldfish. Eggs obtained from four females were fertilized with semen obtained from four males and were subsequently incubated at 18-19°C. After hatching, the free embryos were transferred to experimental rearing units and acclimated to the temperature of 25°C (for 1 day). Such a thermal regime has been recommended as optimal for both chub eggs and larvae (Kujawa, 2004; Kupren et al., 2011a; 2011b). Larvae were reared in three 15 L aquariums in a closed water system as described by Krejszeff et al. (2010). The initial stocking density of larvae was 85 ind. L<sup>-1</sup>. Fish were exposed to a 12L:12D photoperiod and fed three times a day ad libitum with freshly-hatched Artemia sp. nauplii (San Francisco origin) (Demény et al., 2012). During the larval period, oxygen and pH were maintained at >85%, and 7.7, respectively. Ammonia and nitrite (monitored with a photometer, HI 83200, Hanna Instruments, Italy) were always kept below 0.01 mg L<sup>-1</sup> (Nowosad *et al.*, 2013).

The developmental study was conducted on samples of 20 specimens randomly taken every day from hatching to 30 DPH (Days Post Hatch). The sampled specimens were subjected to over exposure in an anesthetic solution (MS-222 at a dose of 150 mg L<sup>-1</sup>) and digital photographs of each specimen were taken using DP-Soft digital image analysis software (Olympus, Japan). After this, the larvae were fixed in a 4% phosphate-buffered formaldehyde solution for further detailed examinations. For monitoring of the early development of chub during the free embryo and larval phase from hatching loss of larval finfold and acquisition of the full finray complement and beginning of squamation (Kendall et al., 1984), the system of developmental stages proposed for carp Cyprinus carpio L. by Peňaz et al. (1983) was used.

The system was based on the instantaneous stages of ontogeny, i.e. on the characteristic moments of identification development. During developmental stages of chub, special attention was also paid to the flexion of the notochord - one of the key events in the development of many fish, which is important from the point of view of other terminologies of early development (Kendall et al., 1984). Moreover, the onset of metamorphosis was additionally defined using the inflection points of allometric as indicators (Kendall et al., 1984; Mihelakakis et al., 2005). Thresholds during early development, such as the filling of the swim bladder exogenous feeding commencement, considered to be achieved when at least 50% of the specimens represented this particular stage. In each digital photograph, several body proportions associated with feeding and locomotion were measured ( $\pm 0.01$  mm) (Table 1). The body characteristics were measured along or perpendicular to the body axis. Dead or abnormal specimens (with malformations) were excluded from the analysis.

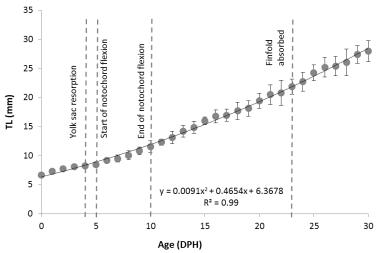
The allometric growth of each character was expressed as a power function of TL, with the intercept and exponent obtained from linear regressions on log-transformed data. When isometric growth occurred, b=1, allometric growth was positive when b>1 and negative when b<1. The inflexion points of growth curves were calculated according to van Snik *et al.* (1997) and Kupren *et al.* (2014).

#### **Results**

Between the day of hatching and 30 DPH, the TL of the European chub body (TL) grew continuously and increased by 21.31 mm (mean length was 6.68 and 27.99 mm in 0 DPH and 30 DPH, respectively) (Figure 1). During this period, twelve developmental stages were observed (ES8 $_b$ -JS1 $_a$ ) (Table 2). The hatching occurred at the stage described as ES8 $_b$ . The yolk sac was completely consumed at 4 DPH (LS2/LS3 $_a$ ), when TL was 8.25±0.27 mm. The notochord flexion occurred between 8.45±0.32 mm TL at 5 DPH (LS3 $_b$ ) and 11.57±1.01 mm TL on 10 DPH (LS5 $_a$ ). The finfold was completely absorbed at 22 DPH (JS1; 20.84±2.15

<b>Table 1.</b> Abbreviations and	description of	morphometric characters	measured in the present study

Character	Abbrev.	Description
Body depth	BD	Posterior to the gill cover
Body depth at anus level	BDA	Posterior to the anus
Eye diameter	ED	Parallel to the longitudinal axis of the body
Head depth	HD	From the bottom of the mouth cavity to the top of the head
Head length	HL	From tip of snout to the margin of gill cover
Snout length	SNL	From the tip of the snout to the eye
Tail depth	TD	Minimum depth of the caudal peduncle
Tail length	TAL	Post anal length
Total length	TL	From the tip of the snout to the end of the caudal fin
Trunk length	TRL	From the end of the operculum to the anus



**Figure 1.** Changes in total length (TL) *L. cephalus* (L.) during early stages of development (from hatching to 30 DPH, days post hath) at 25°C. Bars indicated SD (standard deviation).

mm TL). During the first two days after emergence, the free embryos lay on the bottom of the tank, showed photophobia and tended to congregate in the corners of the tanks. During this time, respiration was accomplished mainly through the body surface and the network of vessels present on the yolk sac and ventral fin fold. Two days after hatching, most of the embryos started to swim up to the surface in order to fill their swim bladders. At 3 DPH, the second chamber of the swim bladder was filled by air and most larvae actively swam and maintained their position in the water column. From that time until the end of the experiment, the larvae were visual feeders, actively searching for food in all parts of the tank.

Table 2 gives details of the stages of free embryo and larval which are illustrated in Figure 2.

The allometric growth patterns of 10 body parts (including TL) were measured in 563 L. cephalus larvae (Figure 3 a-i). Two growth stanzas (biphasic growth) were observed for all measured parameters except for the TRL, which showed negatively isometric growth throughout the studied period (simple linear regression), with a constant growth rate of 0.74 (negatively allometrically) (Figure 3i). In every case, there was a reduction in the growth coefficients after the inflection point. The b value of these body parts differed significantly between their sequential stages (P<0.001). Three parameters: SNL, TAL and BDA showed still positive allometric growth after the inflexion point. For HL, HD and BD isometric growth after the inflexion point were observed. ED changed to negative allometric growth Figure 3 a-h). During the flexion of the notochord (corresponding to the TL interval of 8.45 to 11.57 mm) there was a decline in the growth rate of the tail and head lengths (b = 1.47 to 1.19 and 1.59 to 0.99, respectively). A few days later at the 15.02 mm TL growth of ED and HD slows down to a rate of 0.83 and 1.06 respectively (P<0.001). During the next stages of development, growth concerned all body segments, but was particularly fast for the SNL, BDA and TD. The inflexion points of these parameters were situated at a TL of 24.30, 22.11 and 25.69, respectively (Figure 3).

## **Discussion**

The morphological changes and the size distribution during each developmental stage reported in this study are in accordance with that previously described by Economou et al. (1990) and Çalta (2000) in reared larvae of the same species. In our study, the morphological findings on early development were reported more details and the morphologic findings were supported by allometric growth patterns. At hatching, most functional systems of European chub embryos are incompletely differentiated. Consequently, significant morphogenesis occurs during free embryo and larval development. The newly-hatched individuals have an elongated yolk sac containing the vitelline reserves. The head is bent downwards, the primordial marginal finfold is not differentiated, the eyes lack guanidine pigmentation and the mouth is not formed. During this time, respiration is accomplished mainly through the body surface and the network of vessels present on the yolk sac and ventral finfold. A similar level of development at hatching is characteristic for other lithophilic cyprinids, such as barbus Barbus barbusor nase Chondrostoma nasus (L.), which are not able to swim freely soon after emergence and stay close to the bottom before inflation of the swim bladders (Peňaz, 1974; Krupka, 1988). The level of ontogenic development at hatching is species-specific and highly dependent on environmental conditions, especially of water temperature (Kamler, 1992; Teletchea et al., 2009; Kupren et al., 2011b, Kucharczyk et al., 2014, Palińska et al., 2014). For

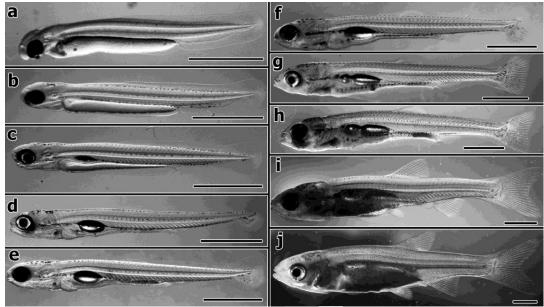
**Table 2.** Developmental stages of European chub *Leuciscus cephalus*(L.) reared at 25°C (ES – embryonic stage; LS – larval stage; JS – juvenile stage; DPH – days post hatch)

Developmental stages	DPH	Total length range, mean (mm)	Characteristic feature	Behaviour
ES8 <sub>b</sub>	0	6.34–7.13 (6.68)	The body is straightened The head bent forward over the yolk sac, but separated, yolk sac pear-shaped. The urostyle is straight. No pigmentation is present on the body, except for the eyes. Ducti Cuvieri visible. Blood circulate from the dorsal aorta to the caudal artery, which supply the caudal vein. The caudal vein consists of a very thin network of twisted vessels (Figure 2a).	Hatching. The embryos show photophobia, lay on the bottom and tend to congregate in the corners of tank.
ES9 <sub>a</sub>	1	6.99–7.56 (7.56)	Reflective guanin in the eyes. Head slightly bent down, body straight. Blood circulation in the branchial arteries. The lower jaw is not formed (mouth is closed). The first stellar melanophores appear on the surface of the body (Figure 2b).	The embryos still show photophobia and lay on the bottom. The embryos cannot swim actively, but short periods of horizontal darting movements are observed.
ES9 <sub>b</sub>	2	7.21–8.17 (7.75)	The whole body is straight. The yolk sac at an advanced stage of resorption. Lower jaw is mobile and reaches the front edge of the eye. The second chamber of the swim bladder is filled by water. The Cuvier ducts have disappeared. The gill covers are developing. The number and size of melanophores on the surface of the body is increasing (Figure 2c).	Embryos start to swim up to the surface in order to fill their swim bladder.
ES9c/ LS1	3	7.59–8.39 (8.14)	The filaments form on the branchial arches. The second chamber of the swim bladder is filled by air. The mouth is in terminal position. Beginning of exogenous (mixed) feeding, the remains of the yolk sac are present (Figure 2d).	Larvae swim actively in the water column.
$LS2/LS3_a$	4	7.73–8.89 (8.25)	The yolk sac is completely resorbed. Lepidotrichia in the caudal and pectoral fin.	Larvae swim actively in the whole water column.
LS3 <sub>b</sub>	5	7.87–8.95 (8.45)	Start of notochord flexion, heterocercal shape of caudal fin (Figure 2e).	Larvae swim actively in the whole water column.
LS4 <sub>a</sub>	7	7.99–10.61 (9.49)	Ossified rays in the caudal fin. Lepidotrichia in the dorsal fin (Fig. 2f).	Larvae swim actively in the whole water column.
LS4 <sub>b</sub>	9	9.42–12.07 (10.84)	The first chamber of swimbladder is filled with gas. Lepidotrichia in the anal fin. The caudal fin is bifurcate and has a definite homocercal shape (Figure 2g).	Larvae swim actively in the whole water column.
LS5 <sub>a</sub>	10	9.04 – 13.13 (11.57)	Notochord flexion finished, pelvic fin buds present, the dorsal fin is almost fully detached from the finfold (Figure 2h).	Larvae swim actively in the whole water column.
LS5 <sub>b</sub>	16	14.44–18.37 (16.77)	Pelvic fins reach the edge of the finfold (Figure 2i).	Larvae swim actively in the whole water column.
LS6	17	15.61–19.23 (16.84)	The pelvic fins extend beyond the edge of the finfold. Finfold present only between the pelvic and anal fin.	Larvae swim actively in the whole water column.
JS1 <sub>a</sub>	22	18.12–22.09 (20.84)	Finfold completely reduced. Scale primordia present (Figure 2j).	Juveniles swim actively in the whole water column.

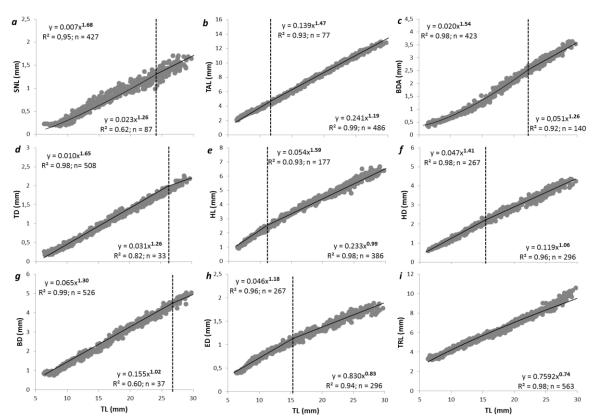
example, many marine species from warm and temperate regions which produce numerous small eggs are extremely poorly developed at hatching (Gisbert *et al.*, 2002; Peña and Dumas, 2009; Çoban *et al.*, 2012).

The ontogenetic shifts in morphology and phenotypic plasticity in specific environments and rearing conditions are reflected in allometry (Simonovic *et al.*, 1999; Khemis *et al.*, 2013).

Teleostean embryonic and larval stages are usually characterized by a high degree of allometric growth patterns (Fuiman, 1983; Osse *et al.*, 2004; van Snik *et al.*, 1997). In juvenile and adult stages, all growth coefficients approach 1 (near-isometric growth). It has been generally accepted that this faster growth is related to rapid development of essential feeding structures (head region), and the need for improved swimming performance (tail region) (Fuiman, 1983;



**Figure 2.** Morphological development of the European chub, *L. cephalus*(L.) (a – stage ES8<sub>b</sub>, 0 DAH; b – stage ES9<sub>a</sub>, 1 DPH; c – stage ES9<sub>b</sub>, 2 DPH; d – stage ES9<sub>c</sub>/LS1, 3 DPH; e – stage LS3<sub>b</sub>, 5 DPH; f – stage LS4<sub>a</sub>, 7 DPH; g – stage LS4<sub>b</sub>, 9 DPH; h – stage LS5<sub>a</sub>, 10 DPH; i – LS5<sub>b</sub>, 16 DPH; j – stage LSJ1<sub>a</sub>, 22 DPH). Scale bars = 2 mm.



**Figure 3 a-i.** Allometric growth equations and relationship between measured body proportions and total length during European chub *L. cephalus* (L.) development (from hatching up to day 30). Morphometric abbreviations are listed in Table 1.

van Snik et al., 1997; Osse and van der Boogart, 2004). In agreement with the former hypothesis, the present results showed that the anterior and posterior body regions of chub specimens initially showed

positive allometric growth. The rapid growth in the head region (especially rapidly before the end of the flexion of the notochord) of chub individuals (allometric growth of HL, HD, ED and SNL), is

probably linked to the development of nervous (midbrain and hindbrain), sensory (vision and olfaction), respiratory (gill arches and filaments) and feeding systems, as increasing head size is associated with a more developed nervous system, allowing better oxygen uptake and the uptake of food particles of increasing size (Fuiman, 1983; van Snik et al., 1997; Koumoundouros et al., 1999; Gisbert et al., 2002; Kupren et al., 2014a, 2014b). In the posterior part of the body, rapid tail growth (allometric growth of TAL, BDA and TD) was concomitant with the development of musculature, unpaired fins, caudal peduncle and fin rays. Such changes contribute to swimming ability improved (increasing probability of zooplankton detection and predator avoidance) (Fuiman, 1983; Gisbert, 1999; Gisbert et al., 2002)

The replacement of temporary organs and the appearance of definitive structures in chub individuals at approximately 21 mm TL did not coincide with a full reduction of relative growth. This particularly relates to BD, BDA, SNL and TD. This pattern of development resulted in the transition from a longitudinally-elongated body shape to more fusiform streamlined body and has been explained as an adaptation to reduce and optimize the energy costs of larval transport (van Snik et al., 1997), reduce drag forces on the body and attain higher locomotion speed. A similar growth tendency is also observed in other fish species with good swimming abilities during their larval period (Koumoundouros et al., 1999; Peña and Dumas, 2009; Kupren et al., 2014b). The positive allometry of body depth (BD) observed during the larval period is also probably linked with the development of the digestive tract and associated glands. In a closely-related species Leuciscus idus (L.), the development of mucosa folds and the coiling of the digestive tract and an increase in enzymatic activity enhance the digestive capability taking place during this time (Ostaszewska et al., 2003)

In many fish species a reduction in the growth coefficients of various organs and tissues occurs at very similar body length sand are correlated to typical morphogenetic events (usually taking place close to the end of the notochord flexion). In Cyprinus carpio Linnaeus, 1758 (van Snik et al., 1997), for example, it was found around 7 mm TL and is probably related to a change in swimming mode from an anguilliform larval-type to a sub-carangi form juvenile-type. The present study showed that the reduction in the growth coefficients of various body parts in chub occurred across a wide range of body lengths and ages (Figure 3). These gradual changes in body proportions are probably related to the lack of sharp changes in behavior and habitat and may be also considered to be a gradual adaptation to life in flowing water. According to the available literature, such changes strictly depend on both environmental (rearing conditions) and genetic factors.

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## References

- Banarescu, P. 1992. Distribution and dispersal of fresh water animals in North America and Eurasia. In: P. Banarescu (Ed.), Zoogeography of Fresh Waters, vol. 2, Aula Verlag, Wiesbaden: 524-1087.
- Bolland, J.D., Cowx, I.G. and Lucas, M.C. 2008. Movements and habitat use of wild and stocked juvenile chub, *Leuciscus cephalus* (L.). Journal of Fish Biology, 71: 1810-1819. doi: 10.1111/j.1095-8649.2009.02244.x
- Çalta, M. 2000. Morphological development and growth of chub (*Leuciscus cephalus* L.) larvae. Journal of Applied Ichthyology, 16: 83-85. doi: 10.1046/j.1439-0426.2000.00164.x
- Choo, CK. and Liew, H.C. 2006. Morphological development and allometric growth patterns in the juvenile seahorse Hippocampus kuda Bleeker. Journal of Fish Biology, 69: 426-445. doi: 10.1111/j.1095-8649.2006.01105.x
- Çoban, D., Cüneyt, S., Şükrü, Y., Şahin, S. and Kürşat, F. 2012. Morphological development and allometric growth of sharpsnout sea bream (*Diplodus puntazzo*) larvae. Turkish Journal of Fisheries and Aquatic Sciences, 12: 883-891. doi: 10.4194/1303-2712v12\_4\_16
- Demény, T., Trenovszki, M.M., Sokoray-Varga, S., Hegyi, A., Urbányi, B., Żarski, D., Ács, B., Miljanović, B., Specziár, A. and Müller, T. 2012. Relative Efficiencies of *Artemia nauplii*, d and Mixed Food Diets in İntensive Rearing of Larval Crucian Carp (*Carassius carassius* L.). Turkish Journal of Fisheries and Aquatic Sciences, 12: 691-698. doi: 0.4194/1303-2712-v12\_3\_18
- Durand, J.D., Unlü, E., Doadrio, I. and Pipoyan, S. 2000.

  Origin, radiation, dispersion and allopatric hybridization in the chub *Leuciscus cephalus*.

  Proceedings of the Royal Society of London B, 267: 1687-1697.
- Economou, A.N., Daoulas, C.H. and Psarras, T. 1991. Growth and morphological development of chub, *Leuciscus cephalus* L. during the first year of life. Journal of Fish Biology, 39: 398-408. doi:10.1111/j.1095-8649.1991.tb04371.x
- Fredrich, F., Ohmann, S., Curio, B. and Kirschbaum, F. 2003. Spawning migrations of the chub in the River Spree, Germany. Journal of Fish Biology, 63: 710-723. doi: 10.1046/j.1095-8649.2003.00184.x
- Freyhof, J., Lieckfeldt, D., Pitra, C. and Ludwig, A. 2005. Molecules and morphology: evidence for introgression of mitochondrial DNA in Dalmatian cyprinids. Molecular Phylogenetics and Evolution, 37: 347-354. doi: 10.1016/j.ympev.2005.07.018
- Fuiman, L.A. 1983. Growth gradients in fish larvae. Journal of Fish Biology, 23: 117-123. doi: 10.1111/j.1095-8649.1983.tb02886.x

- Fukuhara, O. 1986. Morphological and functional development of Japanese flounder in early life stages. Bulletin of the Japanese Society of Scientific Fisheries, 52: 81-91.
- Gisbert, E. 1999. Early development and allometric growth patterns in Siberian sturgeon and their ecological significance. Journal of Fish Biology, 54: 852-862. doi:10.1111/j.1095-8649.1999.tb02037.x
- Gisbert, E., Merino, G.E., Mugeut, J.B., Bush, D., Piedrahita, R.H. and Conklin, D.E. 2002. Morphological development and allometric growth patterns in hatchery-reared California halibut larvae. Journal of Fish Biology, 61: 1217-1229. doi: 10.1111/j.1095-8649.2002.tb02466.x
- Gomułka, P., Żarski, D., Kucharczyk, D., Kupren, K., Krejszeff, S. and Targońska, K. 2011. Acute ammonia toxicity during early ontogeny of chub, *Leuciscus* cephalus (Cyprinidae) Aquatic Living Resources, 24: 211-217. doi: 10.1051/alr/2011111
- Hajkova, K., Pulkrabova, J., Hajslova, J., Randak, T. and Zlabek, V. 2007. Chub (*Leuciscus cephalus*) as a bio indicator of contamination of the Veltava River by synthetic musk fragrances. Archives of Environmental Contamination and Toxicology, 53: 390-396. doi:10.1007/s00244-006-0190-4
- Kamler, E. 1992. Early life history of fish: An energetics approach. Chapman and Hall, Fish and Fisheries Series, 4: London, 167 pp.
- Kendall, A.W., Ahlstrom, E.H. and Moser, H.G. 1984.
  Early life history stages of fishes and their characters.
  In: H.G. Moser, W.J. Richards, D.M. Cohen, M.P.
  Fahay, A.W. Kendall and S.L. Richardson (Eds.),
  Ontogeny and systematics of fishes: American
  Society of Ichthyologists and Herpetologists, Special
  Publication No: 1. Allen Press Inc., Lawrence, KS:
  11-22
- Khemis, B.I., Gisbert, E., Alcaraz, C., Zouiten, D., Besbes, R., Zouiten, A., Masmoudi, A.S. and Cahu, C. 2013. Allometric growth patterns and development in larvae and juveniles of thick-lipped grey mullet *Chelon labrosus* reared in mesocosm conditions. Aquaculture Research, 44: 1872-1888. doi: 10.1111/j.1365-2109.2012.03192.x
- Kirtiklis, L., Grzymkowska, M. and Boroń, A. 2013. Characterization of three species from the subfamily Leuciscinae (Pisces, Cyprinidae) using the nuclear ITS-1 rDNA spacer. Folia Biologica (Krakow), 61: 53-57. doi:10.3409/fb61\_1-2.53
- Koblickaya, A.F. 1981. Key for Identifying Young Freshwater Fishes. Food Industry Publishing House, Moscow, 208 pp. (In Russian).
- Koumoundouros, G., Divanach, P. and Kentouri, M. 1999. Ontogeny and allometric plasticity of *Dentex dentex* (Osteichthyes: sparidae) in rearing conditions. Marine Biology, 135: 561-572. doi: 10.1007/s002270050657
- Krejszeff, S., Kucharczyk, D., Kupren, K., Targońska, K., Mamcarz, A., Kujawa, R., Kaczkowski, Z. and Ratajski, S. 2008. Reproduction of chub *Leuciscus* cephalus L., under controlled conditions. Aquaculture Research, 39: 907-912. doi: 10.1111/j.1365-2109. 2008.01942.x
- Krejszeff, S., Żarski, D.,Kucharczyk, D., Kupren, K., Targońska, K. and Mamcarz, A. 2010. An experimental device for egg incubation and fish larvae rearing under laboratory conditions. Polish Journal of Animal Sciences, 25: 190-199.

- doi: 10.2478/v10020-010-0016-8
- Krupka, I. 1988. Early development of the barbel (*Barbus barbus* Linnaeus 1758). Prace Ústavu Rybárstvi a Hydrobiologie (Bratislava), 6: 115-138.
- Kryzhanovskij, S.G. 1949. Eco-morphological principles of development in carps, loaches and catfishes. Tr. Inst. Morfologii Zhivotnykh, 1: 5-332 (In Russian).
- Kucharczyk, D., Zarski, D, Targońska, K., Łuczyński, M.J., Szczerbowski, A., Nowosad, J., Kujawa, R. and Mamcarz, A. 2014. Inducedartificialandrogenesis in commontench, *Tinca tinca* (L.), usingcommoncarp and common bream eggs. Italian Journal of Animal Science, 13(1): 196-200. doi: 10.4081/ijas.2014.2890
- Kujawa, R.J. 2004. Biological background for rearing reophylic cyprinid larvae under controlled conditions. Dissertations and Monographs, 88, Wyd. UWM, Olsztyn (In Polish, English summary).
- Kupren, K., Mamcarz, A., Kucharczyk, D. and Prusińska, M. 2008. Changes in morphometric parameters in selected early stages of three fish species from the genus *Leuciscus* (Teleostei, Cyprinidae). Archives of Polish Fisheries, 16: 421-436.
- Kupren, K., Żarski, D., Krejszeff, S., Kucharczyk, D. and Targońska, K. 2011a. The effect of stocking density on growth, survival and development of asp *Aspius* aspius (L.), ide *Leuciscus idus* (L.) and chub *Leuciscus cephalus* (L.) larvae during initial rearing under laboratory conditions. Italian Journal of Animal Sciences, 10:e34: 178–184. doi: http://dx.doi.org/10.4081/ijas.2011.e34
- Kupren, K., Mamcarz, A. and Kucharczyk, D. 2011b. Effect of variable and constant thermal conditions on embryonic and early larval development of fish from the genus *Leuciscus* (Cyprinidae, Teleostei). Czech Journal of Animal Sciences, 56: 70-80.
- Kupren, K., Trabska, I., Żarski, D., Krejszeff, S., Palinska-Żarska, K. and Kucharczyk, D. 2014a. Early development and allometric growth patterns in burbot *Lota lota* L. Aquaculture International, 22: 29-39. doi: 10.1007/s10499-013-9680-3
- Kupren, K., Prusińska, M., Żarski, D., Krejszeff, S. and Kucharczyk, D. 2014b. Early development and allometric growth in *Nannacara anomala* Regan, 1905 (Perciformes: Cichlidae) under laboratory conditions. Neotropical Ichtyology, 12: 659–665. doi 10.1590/1982-0224-20130104
- Mann, R.H.K. 1996. Environmental requirements of European non-salmonid fish in rivers. Hydrobiologia, 323: 223-235. doi: 10.1007/BF00007848
- Mihelakakis, A., Tsoklas, C. and Yoshimatsu, T. 2005. Early development of laboratory-reared common dentex, *Dentex dentex* (L). Aquaculture Sciences, 53: 367-376.
- Nikolioudakis, N., Koumoundouros, G., Kiparissis, S. and Somarakis, S. 2010. Defining length-atmetamorphosis in fishes: amulti-character approach. Marine Biology, 157: 991-1001. doi: 10.1007/s00227-009-1379-7
- Nowosad, J., Żarski, D., Biłas, M., Dryl, K. and Kucharczyk, D. 2013. Dynamics of ammonia excretion in juvenile common tench, *Tinca tinca* (L.), during intensive rearing under controlled conditions. Aquaculture International, 21(3): 629-637. doi:10.1007/s10499-012-9596-3
- Nowosad, J., Targońska, K., Chwaluczyk, R., Kaszubowski, R. and Kucharczyk, D. 2014. Effect of temperature on the effectiveness of artificial reproduction of dace

- [Cyprinidae (*Leuciscusleuciscus* (L.))] under laboratory and field conditions. Journal of Thermal Biology, 45: 62-68. doi: 10.1016/j.jtherbio.2014.07.011
- Osse, J.W.M. and van den Boogart, J.G.M. 2004. Allometric growth in fish larvae: timing and function. American Fisheries Society Symposium, 40: 167-194.
- Ostaszewska, T., Wegner, A. and Wegiel, M. 2003. Development of the digestive tract of ide *Leucis cusidus* (L.) during the larval stage. Archives of Polish Fisheries, 11: 181-195.
- Palińska-Zarska, K., Zarski, D., Krejszeff, S., Nowosad, J., Biłas, M., Trejchel, K. and Kucharczyk, D. 2014. Dynamics of yolk sac and oil droplet utilization and behavioural aspects of swim bladder inflation in burbot, *Lota lota* L., larvae during the first days of life, under laboratory conditions. Aquaculture International, 22(1): 13-27. doi: 10.1007/s10499-013-9663-4
- Peña, R. and Dumas, S. 2009. Development and allometric growth patterns during early larval stages of the spotted sand bass *Paralabrax maculato fasciatus* (Percoidei: Serranidae). Scientia Marina, 73(1): 183-189. doi: 10.3989/scimar.2009.73s1183
- Peňaz, M. 1974. Influence of water temperature on incubation and hatching in *Chondro stomanasus* (Linnaeus, 1758). Zoologicke Listy, 23: 53-59.
- Peňaz, M., Prokeš, M., Kouřil, J. and Hamackowa, J. 1983.

- Early development of the carp, *Cyprinus carpio*. Acta Scientiarum Naturalium Brno, 17: 1-39.
- Simonovic, P.D., Garner, P., Eastwood, E.A., Kovac, V. and Copp, G.H. 1999. Correspondence between ontogenetic shifts in morphology and habitat use in minnow *Phoxinus phoxinus*. Developments in Environmental Biology of Fishes, 56: 117-128. doi: 10.1007/978-94-017-3678-7\_9
- Targońska, K. and Kucharczyk, D. 2011. The application of hCG, CPH and Ovopel in successful artificial reproduction of goldfish (*Carassius auratus auratus*) under controlled conditions. Reproduction in Domestic Animals, 46: 651-655. doi: 10.1111/j.1439-0531.2010.01723.x.
- Teletchea, F., Fostier, A., Kamler, E., Gardeur, J.N., Le Bail, P.Y., Jalabert, B. and Fontaine, P. 2009. Comparative analysis of reproductive traits in 65 freshwater fish species: application to the domestication of new fish species. Reviews in Fish Biology and Fisheries, 19: 403-430. doi: 10.1007/s11160-008-9102-1
- Urho, L. 2002. Characters of larvae-what are they? Folia Zoologica, 51: 161-186.
- van Snik, G.M.J., van den Boogaart, J.G.M. and Osse, W.M. 1997. Larval growth patterns in *Cyprinus carpio* and *Clarias gariepinus* with attention to the finfold. Journal of Fish Biology, 50: 1339-1352. doi:10.1111/j.1095-8649.1997.tb01657.x