



New Diatom Records for Turkish Freshwater Algal Flora from Lakes Ladik (Samsun, Turkey) and Hazar (Elazığ, Turkey)

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Abstract

The majority of phycological studies in Turkey focus on seasonal changes of phytoplankton and phytobenthos in lentic and lotic systems, while very few studies related to counting diatoms in sediment cores obtained as a result of paleolimnological work have been published. In this paleolimnological study, seventeen (14+3) new diatom records for freshwater algal flora of Turkey were determined on sediment core samples at five different locations collected in July 2006 from Lakes Ladik (Samsun) and Hazar (Elazığ). We recorded seventeen diatom taxa belonging to Naviculales (8), Cymbellales (2), Eunotiales (2), Bacillariales (2), Fragilariales (1), Tabellariales (1) and Aulacoseirales (1) orders from Lakes Ladik (Samsun) and Hazar (Elazığ). All of the reported taxa are new records for freshwater algal flora of Turkey. General information on the morphology and taxonomy of the species are briefly described in this paper, with their illustrations.

Keywords: Lake Ladik, Lake Hazar, sediment core, new record, diatom, Turkey.

Introduction

Ecologically and economically, aquatic ecosystems are important resources and, to this end, a framework for the management and protection of water resources in each main catchment basin across Europe was established in 2000, known as the European Water Framework Directive (Kelly, 2013). Diatoms are a very diverse group of algae (Mann, 1999; Wojtal, 2009) and good indicators in aquatic ecosystems. They occur in a wide variety of environments and show a broad range of tolerance along several gradients of abiotic factors (Blanco, Ector, Huck, Monnier, & Cauchie, 2008; Rimet, 2012).

Ertan and Morkoyunlu (1998) pointed out that Turkey has 906.118 ha of lakes, 18.000 ha of dam lakes and a 145.000-km long water network. Therefore, Turkey has a great potential of algal diversity in inland waters. Also, situated between large continents (Eurasia and Africa), Turkey comprises areas of very diverse geological, climatic and environmental conditions, so it should expect that the diversity of diatoms is high in this region (Solak, Kulikovskiy, Kaleli & Gönülol, 2016).

Historically, the first freshwater diatom study in Turkey was conducted by Ehrenberg (1844) based on the material collected from the Murat and Aras

Rivers. Subsequently, Schröder (1895) collected samples from Anatolia (the Tarsus River in Mersin) and Brunthaler (1903) investigated diatoms from Lake Iznik. The above studies were followed by the study of Lake Van (the largest lake in Turkey) performed by Legler & Krasske (1940) and Gessner (1957). Contrary to the past, today's freshwater algal studies is progressing rapidly in Turkey. However, the total list of the algal flora of Turkey has not yet been completed. A few checklists including the results of the studies of freshwater algal flora of Turkey were published on different dates (Gönülol, Öztürk, & Öztürk, 1996; Aysel, 2005; Şahin, 2005). In addition to these check-lists, many studies on the new records of algae have been done for the last twenty-five years (Aysel, Dural, & Gezerler-Şipal, 1993; Öztürk, Gönülol, & Öztürk, 1995a, Öztürk, Gezerler-Şipal, Güner, Gönülol, & Aysel, 1995b; Şahin, 2000, 2002, 2007, 2009; Apaydın-Yağcı & Turna, 2002; Atıcı, 2002; Baykal *et al.*, 2009; Ongun-Sevindik, Çelik, & Gönülol, 2010, 2011; Bekleyen, Gokot, & Varol, 2011; Özer *et al.*, 2012; Akar & Şahin, 2014; Yüce & Ertan, 2014; Varol & Fucikova, 2015; Varol & Şen, 2016). These studies contribute largely to the determination of the freshwater algal flora of Turkey. Also, the reliable descriptive information was given about the new records in these publications.

One of the aims of the study is to learn about the

past ecology of the lake and to determine the effect of the changes in the fault on this ecology based on the diatom records accumulated on the sediment cores. Another aim of this study which is also the purpose of this paper is to determine whether there is a new diatom record for the Turkish Freshwater Algal Flora. The present study constitutes the first phase of the studies to be prepared in line with these purposes. We think that this study will contribute to the diatom flora of Turkey.

Materials and Methods

Study Area

When selecting the sampling areas in accordance with the purpose of our study, it was taken into consideration that both lakes are located on the East Anatolian Fault (the Master Fault). Therefore, while one of our sampling areas was the Lake Ladik from the Middle Black Sea Region, the other was the Lake Hazar from the Eastern Anatolia Region.

Lake Ladik (40°50'N to 41°00'N, 35°40'E to 36°05'E) is located within the borders of the Ladik district of Samsun Province in the central Black Sea region of Turkey (Figure 1). The lake, approximately 5 km-long and 2 km-wide, has an elevation of 867 m and a depth of seasonally ranging between 2.5 and 6.0 m. Lake Ladik has a drainage area of 141.40 km² (SHW, 1997). In addition to Küpecik, Çakırgümüş and Talıcak inlet streams, the lake is fed by small streams coming from the Akdağ Mountain. Water level of the lake is controlled by the Tersakan stream that flows into Yeşilirmak River. Lake Ladik was classified as an eutrophic lake (Maraslıoğlu, Soylu, & Gönülol, 2005) but it is under natural protected area due to its floating islands (Bulut, 2012). Prior to 1958 when a regulator was constructed at the Tersakan outlet, the natural size of the lake was much smaller (Figure 1). This regulator could only stabilized the

water level of the lake. Following the improvement of the regulator in 1986, the water depth in the lake has increased significantly and has become its present state (SHW, 1997).

Lake Hazar, formed by faulting of the Eastern Anatolia Fault, is a tectonic lake located at 38°31' N, 39°25' E. Lake Hazar covers about 100 km² of the basin area with an NE-trending elongated-shape along the East Anatolian Fault. The Hazar Basin is a 25 km-long, 7 km-wide and 216 m-deep depression located on the central section of the East Anatolian Fault zone at a mean elevation of 1250 m and predominantly overlain by Lake Hazar. This basin has been described previously as a pull-apart basin because of its rhombic shape and an apparent fault step-over between the main fault traces situated at the southwestern and northeastern ends of the lake. According to bathymetrical measurement of the lake, various maximum depths ranging between 80 and 210 m (Figure 2). The bathymetry map shows two distinct areas: an irregular 216 m-deep sub-basin in the northeastern half of the lake and a much flatter and shallower sub-basin occupying its southwestern half. These two main sub-basins have been named the Deep and Flat basins to facilitate the description of the basin. Hazar Basin sediments are predominantly sourced from two main drainage systems discharging into the lake at its southwestern and northeastern ends (Hempton, Dunne, & Dewey, 1983). The northeastern drainage system comprises several streams with small drainage areas, whereas the southwestern system is mainly dominated by the Kürk River, which drains a large area and forms a significant alluvial fan at the southwestern coast of the lake (Figure 2). (Moreno *et al.*, 2011).

Sampling and Identification

Sediments were sampled in both Lakes Ladik and Hazar using a UWITEC gravity corer in July



Figure 1. Location of the sediment core labeled as “LA-001” core reference in Lake Ladik. The gray-shaded area is the approximate natural extend of the lake before it became a water reservoir in 1958.

2006. In Lake Hazar, 84 and 94 cm-long cores were collected from the southwestern edge of the lake at water depths of about 5 meter (HZ002 and HZ003, respectively). The other two cores in Hazar, 74 and 62 cm-long, were obtained at water depths of 20 and 57 meters (HZ005 and HZ006, respectively). In Lake Ladik, LA001 core recovered the top 62 cm of the sedimentary sequence at a water depth of 4 meter. The coordinates, lengths and taken depths of the cores are shown in Table 1. The cores remained refrigerated in dark and at 4-6 °C temperature before sub-sampling in the laboratory prior to analysis.

Diatom valves were separated from the organic matrix of the sediments using standard techniques defined by Batterbee (1986). Small samples of dry sediment (0.5-1 g) were digested in a solution of 50:50 concentrated sulfuric and nitric acid. Digestion was accelerated by placing the samples in a boiling water bath for 1 h. Digested slurries were rinsed several times in distilled water until neutral pH was reached. On a slide warmer, slurries were dried overnight on coverslips, and then mounted onto microscope slides using Naphrax, a permanent mounting medium.

Diatom valves were enumerated and identified under oil immersion (1000X magnification) using an Olympus BX41 microscope. For each slide, a minimum of 300 valves was counted along a transect. Species identifications were based primarily on the Krammer and Lange-Bertalot (1991a, 1991b, 1999a,

1999b). We checked checklist of Aysel (2005) and the database of Turkish algae (Gönüloğlu, 2017), to determine new taxa for Turkish freshwater algal flora. The current accepted nomenclature has been checked with AlgaeBase (Guiry & Guiry, 2017) and the author names are given in abbreviated form according to Brummit and Powel (1992).

Results and Discussion

A total of seventeen (14+3) diatoms: Naviculales (8), Cymbellales (2), Eunotiales (2), Bacillariales (2), Fragilariales (1), Tabellariales (1) from Bacillariophyceae classis and Aulacoseirales (1) from Coscinodiscophyceae classis were listed below.

Divisio: Bacillariophyta

Class: Bacillariophyceae

Order: Naviculales

Family: Brachysiraceae

Genus: *Brachysira* Kützing, 1836

Brachysira brebissonii Ross, 1986 (Figure 3, a)

Synonyms: *Navicula aponina* var. *brachysira* Kütz.

Navicula brachysira Brébisson ex Rabenhorst

Navicula serians var. *minima* Grunow

Navicula serians var. *minor* Grunow

Anomoeoneis brachysira (Brébisson ex

Rabenhorst) Cleve

Schizonema brachysirum (Brébisson) Kuntze

Anomoeoneis serians f. *minor* Boyer

Anomoeoneis serians var. *brachysira* (Brébisson

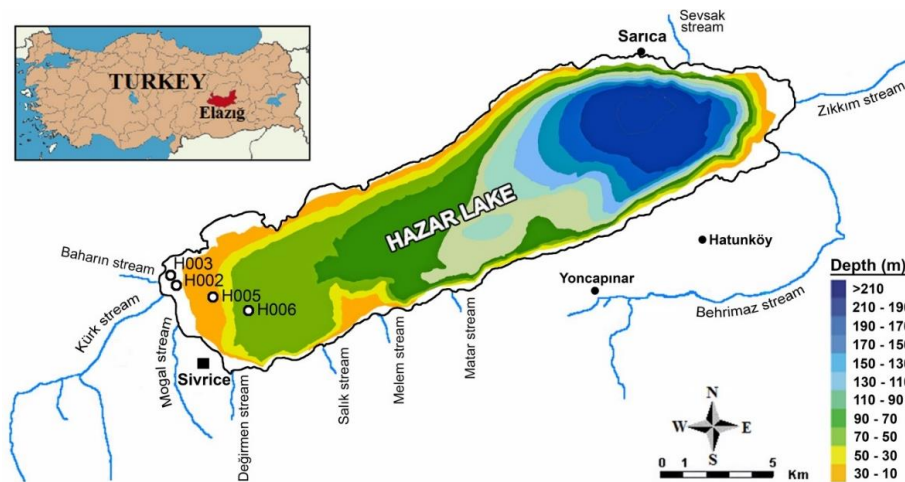


Figure 2. The bathymetry map and sediment core locations of Lake Hazar (H002, H003, H005, H006).

Table 1. The coordinates, lengths and taken depths of the cores in Lakes Hazar and Ladik

Core reference	Coordinate		Depth (m)	Length (cm)	
	(N.)	(E.)			
Hazar	HZ002	38°27'59.64"	39°18'09.73"	5	84
	HZ003	38°28'13.72"	39°18'00.15"	5	92
	HZ005	38°27'41.31"	39°18'39.64"	20	68
	HZ006	37°27'26.73"	39°19'11.18"	57	74
Ladik	LA001	40°53'59.57"	36°01'50.92"	4	62

ex Rabenhorst) Hustedt

Anomoeoneis serians var. *minor* (Grunow)
F.W.Mills

Anomoeoneis brachysira var. *minima* (Grunow)
A.Cleve

Description: Length range: 11-36 µm, width range: 4-8 µm, striae in 10 µm: 26-35. Valves are small, rhombic-lanceolate to elliptical-lanceolate with narrow rounded apices. Striae are weakly radiate from center to apex. The raphe is positioned within two ribs that lie on the external face of the valve (Hamilton, 2010).

Habitat Characteristics: This is a freshwater species and its general habitat is phytoplankton, benthic and fossil. *Brachysira brebissonii* is acidophilous, it has an affinity for environments with pH<7, with an optima of ~ 5.9 (Stevenson, Peterson, Kirschtel, King, & Tuchman, 1991). This taxon was widely distributed across North America in acidic waterbodies. In studies along the Atlantic coastal plain this taxon was observed in ponds that were moderately acidic and oligotrophic to mesotrophic. Despite being widespread, it was usually found in low numbers. pH and TP optima (average weighted means, AWM) from four studies (Siver & Hamilton, 2011; Siver, Hamilton, Stachura-Suchoples, & Kocielek, 2005; Camburn & Charles, 2000; Gaiser & Johansen, 2000) range from 4.7-5.8 and 5.7-13.2 for pH and TP (µg/L) respectively. Also, *Brachysira brebissonii* is the most abundant taxa in dilute boreal and arctic lakes (Wolfe & Kling, 2001). We found this species at H005 and H006 stations of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Finland, Germany, Netherlands, Romania, Russia (Europe), Ireland, Spain, Baltic Sea, Britain, France, Italy, Macedonia, and Poland), *South-west Asia* (Iraq and Israel), *North America* (Northwest Territories, United States of America, Virginia, Alaska, Florida, Georgia, Great Lakes, New Jersey, Wyoming, NW USA and Tennessee), *South America* (Colombia and Brazil), *Asia* (Taiwan, China, Russia (Far East), and Nepal), *Atlantic Islands* (Iceland), *Pacific Islands* (Hawaiian islands), *Africa* (Ghana and Kenya), *Australia and New Zealand* (New Zealand, Tasmania, New South Wales, Northern Territory, Queensland, Western Australia, and Victoria) (Guiry & Guiry, 2017).

Brachysira styriaca (Grunow) R.Ross, 1986 (Figure 3, b)

Basionym: *Navicula styriaca* Grunow

Synonyms: *Navicula styriaca* Grunow

Frustulia styriaca (Grunow) De Toni

Brebissonia styriaca (Grunow) Kuntze

Anomoeoneis styriaca (Grunow) Hust.

Description: Length range: 16-37 µm, width range: 4.9-7.1 µm, striae in 10 µm: 26-30. Valves are lanceolate with rounded, unprotracted apices. Striae are radiate. (Bahls, 2014).

Habitat Characteristics: This is a freshwater species and is widely distributed and locally abundant in northern and alpine regions of the Northern

Hemisphere (Lange-Bertalot & Moser, 1994). This species appears to favour waters with pH readings of <7 (Foged, 1974; Siver *et al.*, 2005). We found this species at H006 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Finland, Germany, Romania, Russia (Europe), Ireland, Spain, Baltic Sea, Britain, France, Italy, Macedonia, and Poland), *North America* (Northwest Territories, Alaska, Great Lakes, NW USA, United States of America), *Africa* (Sudan), *Asia* (Russia (Far East)), *Atlantic Islands* (Iceland), *Pacific Islands* (Hawaiian Islands), *Australia and New Zealand* (New South Wales, Queensland, Victoria, Australia, New Zealand, Tasmania) (Guiry & Guiry, 2017).

Brachysira cymbelliformis Metzeltin and Lange-Bert. 2007

Description: Valves strongly dorsiventral like in *Encyonema* or *Amphora*. Length 12.7-30 µm, breadth 4-7 µm. Raphe straight, filiform. Striae subparallel in central parts becoming progressively radiate towards the apices, 21-22 in 10 µm (Metzeltin & Lange-Bertalot, 2007).

Habitat Characteristics: This is a fossil species seemingly without extant populations. This is a freshwater species. We found this species at H002 in Lake Hazar stated as a lake with oligotrophic conditions by Koçer and Şen (2012). This species has been only found in Florida Peat of the United States by Lange-Bertalot up to now (Metzeltin & Lange-Bertalot, 2007).

Distribution: According to algaebase, there is no information about the bio-geography of this species.

Family: Naviculaceae

Genus: *Caloneis* Cleve, 1894

Caloneis silicula var. *truncata* (Kütz.) Meister, 1912 (Figure 3, c)

Basionym: *Navicula truncata* Kütz.

Synonym: *Caloneis ventricosa* (Ehrenb.) F.Meister var. *minuta* (Grunow) R.M.Patrick

Description: Length range: 25-31 µm, width range: 7-8 µm. Valves are linear and biconstricted, with apices rounded. The raphe is lateral and arched slightly. The striae are radiate to parallel. (Donkin, 2012).

Habitat Characteristics: This is a freshwater species. We found this taxon at H003 and H006 stations of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Netherlands and Romania) and *Asia* (China) (Guiry & Guiry, 2017).

Family: Naviculales incertae sedis

Genus: *Chamaepinnularia* Lange-Bert. and Krammer, 1996

Chamaepinnularia bremensis (Hust.) Lange-Bert., 1996 (Figure 3, d)

Basionym: *Navicula bremensis* Hust.

Description: Valves linear, broadest at the middle with apices rounded. Length 9-12 µm, breadth 2-3 µm. Raphe filiform, with dilated external proximal ends. Striae radiate, not punctate, 18-20/10 µm

(Lange-Bertalot & Metzeltin, 1996).

Habitat Characteristics: This is a freshwater/terrestrial species. We found this species at H002, H003 and H005 stations of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Britain, Germany and Netherlands), *North America* (United States of America and NW USA), *Asia* (Russia (Far East)), *Australia and New Zealand* (New Zealand, Northern Territory and Tasmania) (Guiry & Guiry, 2017).

Family: Naviculaceae

Genus: *Navicula* Bory, 1822

Navicula ammophila Grunow, 1882 (Figure 3, e)

Description: Length range: 27-31 µm, width range: 6.0-6.8 µm, striae in 10 µm: 14-16. Valves are lanceolate to linear-lanceolate with protracted apices. The central area is round. Striae are straight and slightly radiate around the center, becoming parallel, then convergent at the apices (Potapova, 2011).

Habitat Characteristics: This is a freshwater species. We found this species at H005 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Adriatic Sea, Baltic Sea, Germany and Netherlands), *North America* (Mexico, Mississippi and NW USA) and *Africa* (Ghana) (Guiry & Guiry, 2017).

Navicula minima Grunow, 1880 (Figure 3, f)

Synonyms: *Schizonema minimum* (Grunow) Kuntze

Navicula minutissima Grunow

Navicula atomoides Grunow

Navicula tantula Hust.

Description: Length range: 5-18 µm, width range: 2.0-4.5 µm (Krammer & Lange-Bertalot, 1999a). Valves linear to linear-elliptical with bluntly rounded apices. Striae fine, slightly radiate but shorter and more widely spaced at the centre, forming a butterfly-shaped to rectangular central area. Raphe fissures straight, slightly expanded at the centre.

Habitat Characteristics: This is a freshwater species and its general habitat is periphyton, epiphyte and fossil. *Navicula minima* mainly occurs at pH>7 (Alkaliphilous). Its saprobicity is α-meso/polysaprobous in Netherlands. This species is tolerant of moderate pollution and heavy organic pollution. According to TDI (UK), this species mainly occurs between 0.35 and 1.0 mg/l filtrable phosphate (Van Damm, Mertens, & Sinkeldam, 1994; Rott *et al.*, 1999; Kelly *et al.*, 2001). This species was also found by Montoya-Moreno, Sala, Vouilloud, Aguirre, and Plata (2013) in a lagoon. We found this species at H003 and H006 stations of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Baltic Sea, Black Sea, Britain, Czech Republic and/or Slovakia, Finland, Germany, Ireland, Italy, Russia (Europe), Romania, and Spain), *South-west Asia* (Iraq), *Atlantic Islands*, *North America*, *South America*, *Africa*, *Asia*, *Pacific Islands*, *Australia and New Zealand* (Guiry & Guiry, 2017).

Navicula caenosus J.R.Carter and Bailey-Watts, 1981

Description: Valves are 25 µm long and 7 µm broad

with striae 3 in 10 µm (Hartley, Barber, Carter, & Sims, 1996).

Habitat Characteristics: This is a freshwater species. It is reported, a study in Scotland, to be found in lochs (Williams & Reid, 2002). We found this species at H005 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Britain) (Hartley, Ross, & Williams, 1986; Hartley *et al.*, 1996; Whitton, John, Kelly, & Haworth, 2003).

Order: Cymbellales

Family: Cymbellaceae

Genus: *Cymbella* C.Agardh, 1830

Cymbella cymbiformis var. *nonpunctata* Fontell, 1917 (Figure 3, m)

Description: Length range: 25-87 µm, width range: 9-15 µm, dorsal striae in 10 µm: 10-15, ventral striae in 10 µm: 11-16 (Patrick & Reimer, 1975). Large forms found in a study exceeded the length range given above from his taxon (Johansen, Rushforth, Orbendorfer, Fungladda, & Grimes, 1983). The valve of these large forms is often curved transapically and swollen in the midregion.

Habitat Characteristics: This is a freshwater species. According to the our finding it is also a fossil species. We found this taxon at H003 and H006 stations of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Britain, Germany, Russia (Europe), Netherlands and Romania), *South-west Asia* (Iraq), *North America* (Indiana, Iowa, Michigan, Montana, and United States of America), *Asia* (China and Taiwan), *Australia and New Zealand* (Western Australia) (Guiry & Guiry, 2017).

Family: Gomphonemataceae

Genus: *Gomphonema* Ehrenb., 1832

Gomphonema anjae Lange-Bert. and Reichardt, 1991

Description: Valve is 18 µm long and 5 µm broad with striae 7 in 10 µm (Reichardt & Lange-Bertalot, 1991).

Habitat Characteristics: *Gomphonema anjae* is a phytoplanktonic species with freshwater habitat. This species was also found in a lagoon (Colombia) (Montoya-Moreno *et al.*, 2013) and in Animas river (USA), where heavy metals loading is present (Sgro, Poole, & Johansen, 2007). Otherwise, it is a fossil species found in some sediment core samples (Voigt, Gröger, Baier, & Meischner, 2008; Loakes, 2015). We found this species at H003 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Germany and Netherlands), *North America* (NW USA) and *South America* (Colombia) (Guiry & Guiry, 2017).

Order: Eunotiales

Family: Eunotiaceae

Genus: *Eunotia* Ehrenb., 1837

Eunotia soleirolii (Kütz.) Rabenhorst, 1864 (Figure 3, g)

Basionym: *Himantidium soleirolii* Kütz.

Synonyms: *Himantidium soleirolii* Kütz.

Eunotia pectinalis var. *soleirolii* (Kütz.)

H.F.Van Heurck

Description: Length range: 30-65 µm, width range: 3-4 µm, striae in 10 µm: 12-14. Valve linear-lunate in shape. It forms filaments in which the frustules contain internal septae (Ortiz-Lerín & Cambra, 2007).

Habitat Characteristics: *Eunotia soleirolii*, a periphytic algae living in freshwaters, is a fossil species. This species was also found by Montoya-Moreno *et al.* (2013) in a river and a lagoon. We found this species at H005 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Britain, Finland, Germany, Poland, Corsica, Netherlands and Romania), *North America* (New Hampshire, New Jersey, Northwest Territories, NW USA, Tennessee, United States of America), *South America* (Colombia), *Asia* (Russia (Far East) and Korea), *Pacific Islands* (Hawaiian Islands), *Australia and New Zealand* (New Zealand) (Guiry & Guiry, 2017).

Eunotia ambivalens Lange-Bert. and Tagliaventi, 2011 (Figure 3, h)

Synonym: *Eunotia bilunaris* var. *linearis* (Okuno) Lange-Bert. and Nörpel

Description: Length range: 44-205 µm, width range: 3.5-5.5 µm, striae in 10 µm: 9-12. Valves are very slender, elongated with parallel margins and slightly bent to almost straight. Transapical striae is dense and regular. Raphe is in the valve surface with a stroke-shaped appendage (Ortiz-Lerín & Cambra, 2007).

Habitat Characteristics: This is a freshwater species. We found this species at LA-001 station of eutrophic Lake Ladik in Samsun (Turkey).

Distribution: *Europe* (Germany and Netherlands) and *Asia* (Russia (Far East)) (Guiry & Guiry, 2017). According to some records, this species was also found in Bulgarian (Isheva & Ivanov, 2014) from Europe and in some states from North-South America (Bahls, 2009; Montoya-Moreno *et al.*, 2013).

Order: Bacillariales

Family: Bacillariaceae

Genus: *Nitzschia* Hassall, 1845

Nitzschia acula (Kütz.) Hantzsch, 1861 (Figure 3, i)

Basionym: *Synedra acula* Kütz.

Synonyms: *Synedra acus* var. *acula* (Kütz.) Grunow
Nitzschia dissipata var. *acula* (Kütz.) Van Heurck

Description: Valves are 115-235 µm long, 2.8-3.6 µm broad, narrow, linear and needle like, striae 16-18 in 10 µm (Krammer & Lange-Bertalot, 1999b).

Habitat Characteristics: This is a freshwater species, benthic. The species was also found by some researchers in the lake where the pH is low (Camburn & Charles, 2000), but we found this species at H003 station of Lake Hazar stated as an alkaline lake by Koçer and Şen (2012), in Elazığ (Turkey). The optimal TP consumption per year is 63.2 µg/L (Bennion, 1994). Tycho planktonic; Usually found in freshwater and brackish water environment. *Nitzschia acula* markedly prefers less oxygenated waters but higher nutrient concentrations and is dominant in

lentic waters of irrigations channels and marshes. (Pardal, Marques, & Graça, 2002).

Distribution: *Europe* (Baltic Sea, Britain, Germany, Ireland, Macedonia, and Spain), *South-west Asia* (Iran), *North America* (Alaska, Great Lakes, NW USA, Québec, Mexico and United States of America), *Asia* (Mongolia and Taiwan), *Australia and New Zealand* (Queensland and Tasmania) (Guiry & Guiry, 2017).

Nitzschia siliqua Archibald (Figure 3, i)

Synonym: *Nitzschia pseudocarinata* Cholnoky 1970

Description: Valves linear, with margins parallel, tapering quickly to narrow apices with knob-like ends. Length 19-21 µm, breadth 3-4 µm. Keel narrow, and keel puncta indistinct, numbering about the same as the striae, 20-22 in 10 µm (Krammer & Lange-Bertalot, 1988).

Habitat Characteristics: *Nitzschia siliqua* is a rare species. So, habitat information of this species is not available on record. But we found this species at H005 in oligotrophic Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Netherlands), *North America* (NW USA and United States of America), *Australia and New Zealand* (Queensland) (Guiry & Guiry, 2017).

Order: Tabellariales

Family: Tabellariaceae

Genus: *Tabellaria* Ehrenb. ex Kütz., 1844

Tabellaria quadrisepitata B.M.Knudson, 1952 (Figure 3, j)

Description: Length range: 23-130 µm, width range: 6-9 µm. Valve is linear between inflated area and poles. Septa are few (usually 4) (Krammer & Lange-Bertalot, 1991b).

Habitat Characteristics: *Tabellaria quadrisepitata* a freshwater species. This species is mainly occurs at pH<5.5 (Acidobiontic). Saprobicity is oligosaprobous (i.e. oxygen saturation (%)>85, BOD₅<2) in Netherlands. This species is very sensitive to pollution. According to TDI (UK), *T. quadrisepitata* occurs optimum between 0.01 and 0.035 mg/l filtrable phosphate and prefers low nutrient concentrations (Van Damm *et al.*, 1994; Rott *et al.*, 1999; Kelly *et al.*, 2001). We found this species at LA-001 station of eutrophic Lake Ladik in Samsun (Turkey).

Distribution: *Europe* (Belgium, Britain, Germany, and Netherlands), *South-west Asia* (Iraq) and *North America* (Alaska, California, Great Lakes, Minnesota, New Jersey, NW USA, Québec, and United States of America) (Guiry & Guiry, 2017).

Order: Fragilariales

Family: Fragilariaceae

Genus: *Staurosira* Ehrenb., 1843

Staurosira construens var. *exigua* (W.Sm.) H.Kobayasi, 2002 (Figure 3, k)

Basionym: *Triceratium exiguum* W.Sm.

Synonyms: *Triceratium exiguum* W.Sm.

Fragilaria exigua (W.Sm.) Lemmermann

Fragilaria construens var. *exigua* (W.Sm.) Schulz

Description: Length: 9 µm, width: 3 µm, depth: 3 µm

(Krammer & Lange-Bertalot, 1991a).

Habitat Characteristics: This is a freshwater species. We found this taxon at H005 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Britain, Ireland, and Netherlands), *North America* (United States of America), *Australia and New Zealand* (New Zealand) (Guiry & Guiry, 2017).

Class: Coscinodiscophyceae

Order: Aulacoseirales

Family: Aulacoseiraceae

Genus: *Aulacoseira* Thwaites, 1848

Aulacoseira alpigena (Grunow) Krammer, 1991 (Figure 3, 1)

Basionym: *Melosira distans* var. *alpigena* Grunow

Synonyms: *Melosira distans* var. *alpigena* Grunow

Melosira italica var. *alpigena* (Grunow)

A.Cleve

Aulacoseira distans var. *alpigena* (Grunow)

Simonsen

Aulacoseira lirata var. *alpigena* (Grunow)

E.Y.Haworth

Description: Frustules are cylindrical and 4-15 µm in diameter, with a mantle height of 4-7 µm. The valve face is unornamented except for one row of marginal areolae. Spines are located at the end of each perivalvar costa. (Potapova, 2009).

Habitat Characteristics: This is a freshwater phytoplanktonic species. We found this species at H003 station of Lake Hazar in Elazığ (Turkey).

Distribution: *Europe* (Ireland, Netherlands, Spain, Sweden, Britain, Russia (Europe), Baltic Sea, Germany, Ireland, and Romania), *South-west Asia* (Bangladesh and Israel), *Atlantic Islands* (Iceland), *North America* (Alaska, Aleutian Islands, Great Lakes, Northwest Territories, NW USA, Tennessee and United States of America), *South America* (Brazil, Colombia), *Asia* (Mongolia, Korea, Russia (Far East), and Mongolia), *Africa* (Ghana), *Australia and New Zealand* (New Zealand) (Guiry & Guiry, 2017).

The number of diatoms have been estimated to include approximately 10^5 species in over 1,000 genera (Mann, 1999; Fourtanier and Kociolek, 1999, 2003). However, Ichimura (1996) has suggested that the total number of diatom species worldwide is probably not less than 2×10^5 . Diatoms would thus be confirmed as the most species-rich group algae. In total, more than 800 diatom taxa have been found in Turkish lentic and lotic systems to date. The diversity of diatoms in Turkey are relatively low compared to the total number worldwide. The number of diatoms in Turkey are also low compared to other countries, such as Korea (1,000 diatom species) (Park, Lee, Kang, & Lee, 2014). Some genera such as *Navicula*, *Nitzschia*, *Cymbella*, *Surirella*, and *Cyclotella* have the high species diversity in Turkey. Moreover, many of these diatoms frequently reported from water bodies in Turkey are taxa well known in European and Asiatic areas because of seeing commonly there,

too (Solak & Wojtal, 2012). To date, the taxa listed in the results of Ladik and Hazar sediment cores have not been recorded in Turkey (Gönülol *et al.*, 1996; Aysel, 2005; Solak, Ector, Wojtal, Ács, & Morales, 2012; Solak *et al.*, 2016; Ongun-Sevindik *et al.*, 2011).

The trophic structures of the both lakes differ from each other. While Lake Ladik is nutrient-rich eutrophic lake (Maraslıoğlu *et al.*, 2005), Lake Hazar has oligotrophic characters (Koçer & Şen, 2012). So, it is quite normal that the species composition derived from ecological difference of the two lakes is different. Also, when we compare the two lakes in terms of age, it is seen that Lake Hazar is an older lake. Anyway, Moreno *et al.* (2011) stated in a study that Lake Hazar level was ~80 m lower at the start of the Holocene than in 2007 and may have reached its present height only during the last two centuries. This information also confirms that the Lake Hazar is a rather old lake due to being available since the Holocene period. Therefore, it was not too surprising for us that the most of the new diatom records were found in Lake Hazar. While 14+1 new diatom taxa were found in Hazar sediment, 2 new diatom taxa were found in Ladik sediment

According to the algaebase records (Guiry & Guiry, 2017), Bacillariophyceae class constitutes about 80% of Bacillariophyta division and Naviculales order with 4996 species forms the extensive group in the this class. With 1824 species, order Cymbellales forms the second largest group in this class after order Naviculales. This supports the results of which we have found in our study. Among the new diatom records, the Naviculales order from Bacillariophyceae contains the highest (8) records.

Navicula cells are extremely common (in freshwater or marine, epipelagic); hardly a sample can be taken of epipelagic without encountering this genus (Round *et al.*, 1990). The prevalence of *Navicula caenosus* species is not very broad. So, according to the records, the species was previously detected in Houlma Water, Shetland (Williams & Reid, 2002).

Brachysira cells are especially common in oligotrophic lakes and bogs (Round *et al.*, 1990). Freshwater or marine (1 species) to hypersaline; epipelagic. Lange-Bertalot & Moser (1994) report that *B. styriaca* is widely distributed and locally abundant in northern and alpine regions of the Northern Hemisphere and that it is a good indicator of oligotrophic and oligosaprobic waters. We found in line with three new diatom records of the *Brachysira* genus including *B. styriaca* in oligotrophic Lake Hazar. *Brachysira* is a remarkable genus. Although *B. aponica* species was found in marine samples from all world, the other species from *Brachysira* genus were equally widely dispersed but only in freshwater. Some species are abundant in acidic (low conductivity) freshwaters and others are reported from more alkaline sites (Round *et al.*, 1990). Wolfe and Kling (2001) stated that *Brachysira brebissonii* to be

cosmopolitan and acidophilous. pH and TP optima (average weighted means, AWM) from four studies (Siver & Hamilton, 2011; Siver *et al.*, 2005; Camburn & Charles, 2000; Gaiser & Johansen, 2000) range from 4.7-5.8 to 5.7-13.2 for pH and TP ($\mu\text{g/L}$) respectively. But we found this species in Lake Hazar expressed as an alkaline soda lake by Koçer and Şen (2012) due to having the mean pH value of 9.1. Similarly, Kilroy (2007) found that *B. brebissonii* species was abundant in New Zealand subalpine mire pools with alkaline character. This result confirms our findings. *Brachysira brebissonii* is highly polymorphic with respect to valve shape. Wolfe and Kling (2001), in a survey on some *Brachysira* species, noted that *B. brebissonii* has the co-occurrence of lanceolate, elliptical, and more rostrate forms in valve outline. They suggested that the slightly rostrate forms of the species which other authors had applied the subspecific designations of var. or fo. *thermalis*, not be considered distinct from *B. brebissonii*. Otherwise, *B. arctoborealis* is proposed for the rhombic forms that commonly co-occur with *B. brebissonii* and *B. microcephala*. *B. arctoborealis* is distinct from *B. brebissonii* with respect to both valve morphometry and copula structure, and, as an indicator of low pH environments, should be identified as distinct from *B. brebissonii* in ecological and paleolimnological studies.

Nitzschia cells are found from freshwater to marine; usually epipelagic or planktonic. *N. acula* was found by Camburn and Charles (2000) at diatom flora of low-alkalinity lakes in Northeastern United States. However, this species was found in some lake sediments from Lakes Nicholls (Cameron, Tyler, Rose, Hutchinson, & Appleby, 1993) and Hazar with high alkalinity property. According to Pardal *et al.* (2002), *N. acula* prefers less oxygenated waters with higher nutrient concentrations, such as *Cyclotella meneghiniana*, *Melosira varians*, *Navicula phyllepta* taxa and is dominant in lentic waters of irrigations channels and marshes. However, we found this species in oligotrophic Lake Hazar, which was stated by Koçer and Şen (2012) that it has higher oxygenated waters and less nutrient concentrations.

Eunotia is a large genus that comprises around 200 species (VanLandingham, 1968). In this genus although numerous species are restricted to tropical areas, due to their environmental water preferences: low pH and conductivity, the genus has a world-wide distribution (Metzeltin & Lange-Bertalot, 1998; Sala *et al.*, 2002; Diaz-Castro *et al.*, 2003). The occasional cells found in marine fossil deposits dating back to the Eocene are surprising (Round *et al.*, 1990). *Eunotia* is essentially a freshwater diatom genus frequently associated with acidic waters (Sládeček, 1986; Alles *et al.*, 1991; Cameron, 1995; Pierre, 1996) and epiphyton and metaphyton of oligotrophic or dystrophic situations (Patrick & Reimer, 1966; Lange-Bertalot & Metzeltin, 1996). While only one of two new *Eunotia* records was found in oligotrophic Lake

Hazar, the other *Eunotia* record (*E. ambivalens*) was found in Lake Ladik, which has an eutrophic environment. Montaya-Moreno *et al.* (2013) stated that habitat of *Eunotia soleirolii* species is periphyton and fossil. This result confirms our findings that we found this species in sediment core of Lake Hazar. According to Patrick and Reimer (1966), *E. soleirolii* occurs predominantly in periodic water-bodies as well as under alkaline or acidic conditions, high or low electrolyte content. Also, Van Damm *et al.* (1994) stated that this species is circumneutral, mainly occurring at pH values about 7. Patrick and Reimer's result was confirmed by our findings that we found this species in sediment cores of alkaline Lake Hazar. *E. ambivalens* (syn: *E. bilunaris* var. *linearis*) is acidophilus, mainly occurring at pH<7 (Van Damm *et al.*, 1994). However, we found this species in Lake Ladik which was stated as an alkaline lake by Maraşhoğlu *et al.* (2005) due to pH values ranging between 8.0 and 9.0.

Brachysira brebissonii, *B. cymbelliformis*, *Navicula minima*, *Eunotia soleirolii*, *Gomphonema anjae*, and *Cymbella cymbiformis* var. *nonpunctata*, all were found in core sediments of Lake Hazar, are also fossil taxa. *Gomphonema anjae* species from these taxa is found in the phytoplankton of Germany (Ludwig & Schnittler 1996; Täuscher, 2014), Netherlands (Veen, Hof, Kouwets, & Berkhout, 2015) and North-South America Lakes (Bahls, 2009; Montoya-Moreno *et al.*, 2013), and in the phytobenthos of the Lednice Ponds characterized as highly eutrophic because of rich in organic matter, with high calcium and salt contents (Kopp *et al.*, 2012). Also, this species was recorded as a new taxon for Bulgarian algal flora in a study in Boyanska River (Isheva & Ivanov, 2014). Otherwise, in some studies it was found in sediment cores (Voigt *et al.*, 2008; Loakes, 2015). Ludwig and Schnittler (1996) reported that this taxon is also in red list category (endangered species) for Germany. According to the some Turkish records (Pala, 2007; Sıvacı, Barinova, Solak, & Çobanoğlu, 2013), only three of the six fossil taxa are new diatom records for freshwater algal flora of Turkey. In the some studies, it is stated that *C. cymbiformis* var. *nonpunctata* (Figure 3, m) was previously found by Şahin (1998), Sıvacı and Pabuçcu (2007), Baytut and Gönülol (2016). However, none of the previous studies are on sediment cores and in none of them has been found in fossil records. Two of the three studies were made on periphyton and the other one was on phytoplankton. According to algaebase data (Guiry & Guiry, 2017), however, it is a fossil taxon seemingly without extant populations. Besides, most studies in other countries (e.g. Laird Fritz, & Cumming, 1998; Ognjanova-Rumenova, 2001; Tingstad *et al.*, 2011; White, 2012), *C. cymbiformis* var. *nonpunctata* was found in the core sediments. Therefore, the other records found on *C. cymbiformis* var. *nonpunctata* from Turkey should be *C. cymbiformis* or *C. cymbiformis* var. *longa*.

Because these *Cymbella* taxa resemble each other and both species (*C. cymbiformis*, *C. cymbiformis* var. *longa*) were previously recorded by Aysel (2005). So, our opinion is that *C. cymbiformis* var. *nonpunctata* taxon is the first record for the Turkey's recent diatom flora.

Algal flora studies require rigorous and long-term effort. Long-term monitoring and measuring of new records can prevent mistakes in the identification of species (Ongun Sevindik *et al.*, 2010). Besides, when a species is identified the habitat and ecology informations of the species should also be considered in addition to the morphological characteristic information of the species. Furthermore, for accuracy, experts need constantly updated databases.

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